# AFFINITY AND PHOSPHORYLATION CONSTANTS OF A SERIES OF *O,O*-DIALKYL MALAOXONS AND PARAOXONS WITH ACETYLCHOLINESTERASE\*

Y. C. CHIU, † A. R. MAIN and W. C. DAUTERMAN

Departments of Entomology and Biochemistry, North Carolina State University, Raleigh, N.C., U.S.A.

(Received 13 January 1969; accepted 14 March 1969)

Abstract—The affinity constants  $(K_n)$  and the phosphorylation constants  $(k_p)$  were determined for a series of O,O-dialkyl malaoxons and paraoxons in which the phosphorylalkoxy groups were varied from methyl to butyl. The  $k_p$  values of the di-n-alkyl malaoxons and paraoxons ranged from 43 to 67 min<sup>-1</sup> except for di-n-butyl malaoxon which was only 25 min<sup>-1</sup>. Neither chain length nor leaving group appeared to affect  $k_p$  greatly, although there were minor variations.

Members in the paraoxon series bound better in each case than the comparable member of the malaoxon series. The  $K_a$  values were from two to ten times better for the paraoxon series than the comparable member of the malaoxon series. These values imparted the larger bimolecular rate constants to the paraoxon series. With di-i-propyl malaoxon and paraoxon poor binding occurred as well as a dramatic decrease in  $k_p$  (3 min<sup>-1</sup>).

MAIN¹ demonstrated kinetically that complex formation prior to phosphorylation occurred with cholinesterase and organophosphorus inhibitors. A study by Main and Hastings² with a series of carbalkoxy homologs of malaoxon and serum cholinesterase showed that the affinity as well as the phosphorylation rates varied with the modifications in the "leaving group". Studies on the geometric isomers of Phosdrin demonstrated that the phosphorylation rate rather than affinity accounted for the higher inhibitory power of the cis-isomer when compared to the trans-isomer of Phosdrin.³ Chiu and Dauterman,⁴ studying the effect that structural differences in the "leaving group" of the various homologs of malaoxon and acetoxon had on the affinity and the phosphorylation rates, found that both values varied with structural changes. They also found that an  $\alpha$ -carbethoxy group was necessary for good phosphorylation rates.

The terminology of Equation 1 will be employed, where P is the dialkyl phosphoryl group, X is the leaving group, E the enzyme, EPX the reversible complex and EP the phosphorylated enzyme.

$$E + PX \stackrel{K_a}{\rightleftharpoons} EPX \stackrel{k_p}{\rightleftharpoons} EP \tag{1}$$

The rate constants are  $K_a$  and  $k_p$  and  $k_i = k_p/K_a$ .

<sup>\*</sup> This work was supported by United States Public Health Service Grant ES-00044. Paper No. 2802 of the Journal Series of the North Carolina State University Experiment Station, Raleigh, N.C. † Present address: Section of Neurobiology and Behavior, Cornell University, Ithaca, N.Y. 14850.

The present study was undertaken to determine the effect which comparable modification of the phosphorylating or "remaining" group (P) would have on affinity  $(K_a)$  and on rates of phosphorylation  $(k_p)$  and the overall inhibitory power  $(k_t)$ . Two comparably substituted series of phosphoro di-alkoxy inhibitors were used: the  $O_sO$ -dialkyl malaoxons

In the malaoxon series, the leaving group (X) is aliphatic while in the paraoxon series, it is aromatic. However, the  $pK_a$  of the leaving groups were very similar; 6.96 for diethylthiomalate<sup>2</sup> and 7.15 for p-nitrophenol.<sup>5</sup> Consequently, the two series might be expected to offer an indication of the degree of interaction between the leaving and phosphorylating groups in determining affinity and rates of phosphorylation. Similarly, the results would also permit comparison of an aromatic with an aliphatic leaving group.

### METHODS

Enzyme. Bovine erythrocyte acetylcholinesterase (acetylcholine hydrolase EC 3.1.1.7) was obtained from Sigma Chemical Company, St. Louis, Mo., U.S.A. The enzyme solution was prepared by dissolving 500  $\mu$ M units in 50 ml of 10 mM sodium phosphate buffer, adjusted to pH 7·6, and kept in the cold under a drop of toluene. One  $\mu$ M unit of the enzyme hydrolyzed 1  $\mu$ mole of acetylcholine per min at pH 8·0 at 37°. The solution was stable for weeks and was further diluted before use.

Organophosphorus compounds. The O,O-dialkyl S-(1,2-dicarbethoxy)ethyl phosphorothiolates referred to as the dialkyl malaoxons were prepared by heating on the steam bath 2 molar equivalents of trialkyl phosphite with 1 molar equivalent of tetraethyl dithiodisuccinate.<sup>6</sup>, <sup>7</sup>

The time of heating of tetraethyl dithiodisuccinate and the various trialkyl phosphites was 22 hr with trimethyl and triethyl phosphites, 48 hr with the tripropyl phosphites, and 72 hr with tributyl phosphite. The excess phosphites were removed under vacuum and the compounds were run through a molecular still to remove the remaining traces of phosphites and diethyl 2-(alkylthio) succinates. The yields were as follows: Dimethyl malaoxon, 80%, 0.01 mm 83°; diethyl malaoxon, 77%, 0.15 mm, 135°; di-n-propyl malaoxon, 66%, 0.18 mm, 150°; di-i-propyl malaoxon, 47%, 0.15 mm, 140°; di-n-butyl malaoxon, 57%, 0.08 mm, 145°. Distillation temperatures were the minimum necessary to distill the product on a falling film molecular still with 1-cm path. All compounds were further purified by multimolecular adsorption chromatography. The physical properties and trivial names are given in Table 1.

The O,O-dialkyl p-nitrophenyl phosphates referred to as dialkyl paraoxons were prepared and purified according to the method of Dauterman and O'Brien.<sup>11</sup>

Determination of  $K_a$ ,  $k_p$  and  $k_i$ . The values of  $K_a$ ,  $k_p$  and  $k_i$  were determined by using the procedure of Main and Iverson. The rate of the inhibition reaction of the enzyme with various concentrations of inhibitors was expressed by plotting  $\log v$  against t. The slopes were calculated by regression analysis and gave  $2\cdot 3\Delta \log v/\Delta t$ 

	Boiling point		Phosphorus		Refractive index (n <sup>20</sup> <sub>D</sub> )		
Compound	C°	mm		Found <sup>9</sup> %)	Calcu- lated <sup>10</sup>	Found	Dev. (%)
Dimethyl malaoxon	.,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,,					-	
O,O-dimethyl S-(1,2-dicarbethoxy) ethyl phosphorothiolate	83	0.01	9.9	9.3	1.4673	1.4660	+0.09
Diethyl malaoxon							
O,O-diethyl S-(1,2-dicarbethoxy) ethyl phosphorothiolate	135	0.15	9.05	8-45	1.4674	1.4631	+0.29
Di-n-propyl malaoxon							
O,O-di-n-propyl S-(1,2-dicarbethoxy) ethyl phosphorothiolate	150	0.18	8.36	7.94	1.4675	1.4631	+0.30
Di-i-propyl malaoxon							
O,O-di-i-propyl S-(1,2-dicarbethoxy) ethyl phosphorothiolate	140	0.15	8.36	8.33	1.4675	1.4617	+0·40
Di-n-butyl malaoxon							

TABLE 1. PHYSICAL CONSTANTS OF PHOSPHORYLALKOXY HOMOLOGS OF MALAOXON

values and their standard errors. These values were further used to plot the graphs of  $i\Delta t/2\cdot3\Delta\log v$  against (i) according to the equation  $i\Delta t/2\cdot3\Delta\log v=i/k_p+1/k_i$  where  $k_i=k_p/K_a$ . All  $K_a$ ,  $k_p$  and  $k_i$  values were obtained by regression analysis with the IBM 360 computer according to the method of Wilkinson.<sup>13</sup>

0.08

145

7.77

7.49

 $1.4676 \quad 1.4604 \quad +0.49$ 

O,O-di-n-butyl S-(1,2-dicarbethoxy)

ethyl phosphorothiolate

The rates of inhibition was measured at 5°, pH 7·6 in  $10_m$ M sodium phosphate buffer. The residual enzyme activities were determined with 50 ml 3·0 mM acetylcholine at 25° and pH 7·6 on a Radiometer pH-stat.

Table 2. Affinity, phosphorylation and bimolecular constants  $(K_a, k_p \text{ and } k_i)$  for the inhibition of acetylcholinesterase with O,O-dialkyl malaoxons at 5°, pH 7·6. The homolog concentration (i) and the corresponding inhibition velocity (i $\Delta t/2$ ·3 $\Delta \log v$ ) from which  $K_a$ ,  $k_p$  and  $k_i$  were calculated are also given

Dimethyl		Diethyl		Di-n	Di-n-propyl		Di-i-propyl		Di-n-butyl	
	i∆t		i∆t		iΔt		iΔt		iΔt	
i (mM)	2·3Δlog v (μM min)	i (mM)	2·3Δlog v (μM min)		2·3Δlog v (μM min)	i (mM)	2·3Δlog v (μM min)	i (mM)	2·3Δlog v (μM min)	
2·00 1·50 1·00 0·50 0·02 0·001	66·5 58·0 52·0 43·4 37·3 38·6	3·125 2·500 2·000 1·000 0·050 0·010	131 117 108 87 70 73	2·00 1·50 1·00 0·50 0·10 0·002	113 105 96 92 79 77	4·50 3·00 2·50 2·00 1·00 0·50 0·10	4005 3572 3449 3233 2949 2795 2573	0·3125 0·2000 0·1500 0·1250 0·0800 0·0600 0·0005	39·1 34·2 32·4 31·5 30·6 28·9 26·7	
Const $ K_a \text{ (m)} $ $ k_p \text{ (m)} $ $ k_i \text{ (M)} $	M)	2·4 67·0	± 0·15 ± 2·6 × 10 <sup>4</sup>	Diethy 3.6 ± 0 52.0 ± 2 1.4 × 10	·23 4·5 ·1 58·0	± 0·10 ± 1·2 × 10 <sup>4</sup>	Di-i-prop 8·9 ± 0 3·30 ± 0 0·04 × 1	$\begin{array}{c ccccccccccccccccccccccccccccccccccc$		

#### RESULTS

The values of the inhibition velocity and kinetic constants of the dialkyl malaoxons are presented in Table 2, whereas the plot of the inhibition velocity  $i\Delta t/2 \cdot 3\Delta \log v$  vs. the concentration of inhibitor i is presented in Fig. 1. With the malaoxon homologs, the best affinity  $(1/K_a)$  was found with di-n-butyl malaoxon and the poorest affinity with di-i-propyl malaoxon. The phosphorylation rate  $(k_p)$  of the dimethyl compound was the fastest and the di-i-propyl malaoxon was the slowest. However, the  $k_p$  values

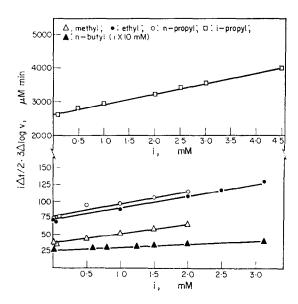


Fig. 1. Inhibition velocity ( $i\Delta t/2 \cdot 3\Delta \log v$ ) vs. concentration (i) for the inhibition of acetylcholinesterase by  $O_iO_j$ -dialkyl homologs of malaoxon.

of the methyl, ethyl and *n*-propyl homologs were of the same order. Overall, the best inhibitor of acetylcholinesterase in the malaoxon series based on the bimolecular rate constant  $(k_i)$  was di-*n*-butyl malaoxon. The average standard errors obtained with the log v vs. t plots at various concentrations of the malaoxon homologs were: Dimethyl  $\pm 2.8\%$ ; diethyl  $\pm 1.6\%$ ; di-*n*-propyl  $\pm 3.0\%$ ; di-*i*-propyl  $\pm 2.3\%$ ; di-*n*-butyl  $\pm 2.2\%$ .

The values of the inhibition velocity and kinetic constants of the dialkyl paraoxons are presented in Table 3 and the plot of the inhibition velocity  $i\Delta t/2 \cdot 3\Delta \log v$  vs. the concentration of the inhibitor i is presented in Fig. 2. Here again the poorest affinity was found with the di-i-propyl paraoxon and the best affinity with the di-n-butyl compound. In the paraoxon series the di-n-butyl compound was the best inhibitor of acetylcholinesterase based on the  $k_i$  value and the di-i-propyl compound the poorest. The average standard errors associated with the paraoxon homologs were: Dimethyl  $\pm 4.9\%$  diethyl  $\pm 1.7\%$ ; di-n-propyl  $\pm 4.9\%$ ; di-i-propyl  $\pm 4.2\%$ ; di-n-butyl  $\pm 3.5\%$ .

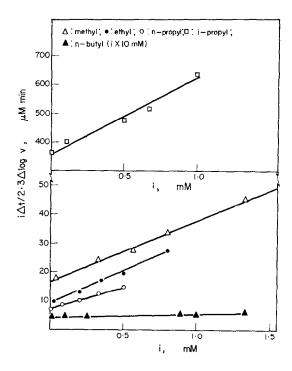


Fig. 2. Inhibition velocity ( $i\Delta t/2\cdot 3\Delta \log v$ ) vs. concentration (i) for the inhibition of acetylcholinesterase by  $O_iO_j$ -dialkyl homologs of paraoxon.

Table 3. Affinity, phosphorylation and bimolecular constants  $(K_a, k_p \text{ and } k_i)$  for the inhibition of acetylcholinesterase with O,O-dialkyl paraoxons at 5°, pH 7·6. The homolog concentration (i) and the corresponding inhibition velocity (i $\Delta t/2\cdot3\Delta\log v$ ) from which  $K_a, k_p$  and  $k_i$  were calculated are also given

Dimethyl		D	Diethyl		Di-n-propyl		Di-i-propyl		Di-n-butyl		
	iΔt		iΔt		iΔt		iΔt		iΔt		
i (m <b>M</b> )	2·3Δlog v (μM min)	i (mM)	2·3Δlog v (μM min)	i (mM)	2·3Δlog v (μM min)	i (mM)	2·3Δlog v (μM min)	i (mM)	2·3Δlog v (μM min)		
2·66 1·33 0·80 0·567 0·333 0·033	69·8 45·4 34·2 27·5 24·5 17·6	0·8 0·5 0·35 0·20 0·01	27·4 19·2 17·2 12·9 9·7	0·500 0·333 0·200 0·0833 0·0033	14·8 12·5 9·6 8·5 6·9	1·000 0·667 0·500 0·100 0·0067	637 515 474 400 364	0·133 0·100 0·089 0·0267 0·0100 0·0065	6·2 5·4 5·5 4·7 4·8 4·7		
Const	Constants		Dimethyl		Diethyl Di-n		-propyl Di- <i>i</i> -pro		oyl Di-n-butyl		
$k_p$ (m	(a (mM) p (min <sup>-1</sup> ) i (M <sup>-1</sup> min <sup>-1</sup> )		50·2 <u>±</u> 1·8 4		$\begin{array}{cccc} 0.36 \pm 0.05 & 0.43 \\ 2.7 & \pm 2.8 & 63.1 \\ 1.20 \times 10^5 & 1.45 \end{array}$		$   \begin{array}{c}     \hline       1.38 \pm 0.2 \\       3.85 \pm 0.2 \\       0.028 \times 1   \end{array} $	27 65·	$\begin{array}{c} 27 \pm 0.06 \\ 0 \pm 11.0 \\ 41 \times 10^5 \end{array}$		

#### DISCUSSION

A comparison of the  $K_a$  values within each of the inhibitor series showed only small variations. In the paraoxon series, the variation between the diethyl, di-n-propyl and di-n-butyl homologs was insignificant suggesting that the acyl-binding site was fully occupied by one or both of the methyl groups of the phosphorylalkoxy portion of the inhibitor. Since  $K_a$  is an equilibrium constant (i.e.  $k_{-1}/k_1$ ) the calculation of the free energy of binding of the inhibitors is justified ( $\Delta F = -RT \ln 1/K_a$ ). In the paraoxon series, going from the dimethyl to the diethyl analog, an increase of about 500 cal/mole in binding energy occurred while in the malaoxon series this change resulted in a decrease of 230 cal/mole. A greater dissimilarity occurred between the di-n-propyl and di-n-butyl malaoxons which was accompanied by an increase in binding energy of 1057 cal/mole. Despite these anomalies, which are minor compared to the potential increase in binding energy expected with the addition of two methylene groups (1000 cal/mole/methylene group), it is seems evident that the n-alkyl substituents at least beyond methyl are not involved in any significant way with initial binding based on the calculated free energy.

In both series, the di-i-propyl substituents resulted in a slightly less binding—about 700 cal/mole in each series. This is probably due to the steric effects of the di-i-propyl group.

Members in the paraoxon series bound better than the comparable member of the malaoxon series. On the average, this difference was of the order of a 1000 cal/mole and accounted almost entirely for the great inhibitory power  $(k_i)$  of the paraoxon series. Since the phosphorylalkoxy portion in the two series is the same, the greater binding capacity must be associated with the *p*-nitrophenoxy group when compared to the diethyl thiomalate moiety.

The phosphorylation rate constants  $(k_p)$  of the di-n-alkyl members were, with one exception, remarkably similar, both between members and between series. The exception was the di-n-butyl member of the malaoxon series which was 25 min<sup>-1</sup> compared with the more typical values which ranged from 42.7 to  $67 \, \text{min}^{-1}$ . Neither leaving group nor chain length appeared to affect  $k_p$  greatly, although there were minor variations. It is perhaps worth noting that the  $k_p$  values were quite high compared with others which have been reported. <sup>15</sup>, <sup>16</sup>

Substitution of the branched-chain di-i-propyl group resulted in a dramatic decrease in  $k_p$  in both series, to about 3 min<sup>-1</sup>, suggesting that the phosphoryl alkoxy substituent does play a major role in determining  $k_p$ . The similar dissociation (or ionizing) constants of the leaving groups suggest a possible correlation with the similar  $k_p$  values,<sup>17</sup> but the similarity could be coincidental, particularly in view of the i-propyl substituent effect.

## REFERENCES

- 1. A. R. MAIN, Science, N.Y. 144, 992 (1964).
- 2. A. R. MAIN and F. L. HASTINGS, Biochem. J. 101, 584 (1966).
- 3. Y. C. CHIU and W. C. DAUTERMAN, Biochem. Pharmac., 18, 359 (1969).
- 4. Y. C. CHIU and W. C. DAUTERMAN, Biochem. Pharmac. 18, 1665 (1969).
- 5. Handbook of Chemistry and Physics, 47th edn., p. D 86. Chemical Rubber Co., Ohio (1966).
- 6. D. E. AILMAN, J. org. Chem. 30, 1074 (1965).
- 7. W. C. DAUTERMAN and A. R. MAIN, Toxic. appl. Pharmac. 9, 408 (1966).
- 8. C. G. PATCHETT and G. H. BATCHELDER, J. Agric. Fd Chem. 9, 395 (1961).

- 9. C. J. BARTON, Analyt. Chem. 20, 1068 (1948).
- 10. R. SAYRE, J. Am. chem. Soc. 80, 5438 (1958).
- 11. W. C. DAUTERMAN and R. D. O'BRIEN, J. Agric. Fd Chem. 12, 318 (1964).
- 12. A. R. MAIN and F. IVERSON, Biochem. J. 100, 525 (1966).
- 13. G. N. WILKINSON, Biochem. J. 80, 324 (1961).
- 14. C. Tonford, in Physical Chemistry of Macromolecules, p. 130. John Wiley, New York (1961).
- 15. A. H. AHARONI and R. D. O'BRIEN, Biochemistry, N.Y. 7, 1538 (1968).
- 16. P. Bracha and R. D. O'BRIEN, Biochemistry N. Y. 7, 1545 (1968).
- 17. T. R. FUKUTO and R. L. METCALF, J. Agric. Fd Chem. 4, 930 (1956).